scientific correspondence

Janual & Court will

be seen exactly what role EMA has in the regulation of E2F-response genes but we suggest that EMA represents the first member of a new class of E2F-like proteins acting to downregulate gene expression in a manner independent of pocket proteins. M. Morkel, J. Wenkel

Laboratory for Molecular Biology of Pediatric Disease, Charité, Humboldt University, Zeigelstrasse 5-9, D-10098 Berlin, Germany

A. J. Bannister, T. Kouzarides

Wellcome/CRC Institute and Department of Pathology, University of Cambridge, Tennis Court Road, Cambridge CB2 IQR, UK

Christian Hagemeier

Department of Paediatrics, Charité-Virchow Medical Center, Humboldt University, Augustenburger Platz 1, D-13353 Berlin, Germany

e-mail: chrish@rz.charite.hu-berlin.de

- 1. Beijersbergen, R. L. & Bernards, R. Biochim, Biophys. Acta 1287, 4 103-120 (1996).
- 2. Weintraub, S. J. et al. Nature 358, 259-261 (1992).
- 3. Weinberg, R. A. Cell 81, 323-330 (1995).
- 4. Chittenden, T. et al. Cell 65, 1073-1082 (1991).
- 5. Kovesdi, I., Reichel, R. & Nevins, J. R. Cell 45, 219-228 (1986).
- 6. Hanna Rose, W. & Hansen, U. Trends Genet. 12, 229-234 (1996).
- 7. Yew, P. R. et al. Genes Dev. 8, 190-202 (1994).
- 8. Weintraub, S. I. et al. Nature 375, 812-815 (1995).
- 9. Hagemeier, C. et al. Nucleic Acids Res. 21, 4998-5004 (1993).
- 10. Voitek, A. B. et al. Cell 74, 205-214 (1993).
- 11. Hagemeier, C. et al. EMBO J. 13, 2897-2903 (1994).

Fluorescence intensity

Figure 2 Induction of apoptosis in KS Y-1 cells by the 18K RNase. KS Y-1 cells (5×105 cells per well, 24well tray) were incubated with purified RNase (a, 4.46 units RNase activity ml-1; b, 1.12 units RNase activity mi⁻¹) in a total volume of 200 μl MDOB medium (16 h, 37 °C) and assayed for apoptosis with the TUNEL reaction (Boehringer-Mannheim). Cells were analysed by FACS; induction of apoptosis (black-enclosed peaks) is shown by an increase in fluorescence intensity attributable to the binding of fluorescent antibody to apoptotic nuclei. Blue peaks are control incubations. Values are percentages of the reading for untreated cells.

Ribonuclease inhibits Kaposi's sarcoma

Kaposi's sarcoma (KS) is a cancer closely associated with AIDS. Crude, commercial human urinary chorionic gonadotrophin (hCG) preparations have previously been found to inhibit the growth of KS cell lines in vitro and in immunodeficient mice1,2, and to induce tumour regression of KS lesions in AIDS patients3,4. Here we report

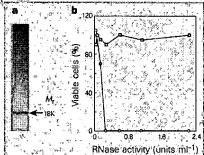


Figure 1 Analysis of RNase from hOG preparations. a, Purified RNase (20 ng protein) was subjected to SDS-PAGE on a Phast gel (gradient 10-15%; Pharmacia Biotech) and the gel stained with silver. b, Effect of the 18K RNase on viability of KS Y-1 (circles) or HeLa cells (squares). Cells (5×10° cells per well, 96-well tray) were incubated with purified RNase (100 µl MDCB medium, Gibco; 16 h; 37 °C) and viability determined with WST-1 reagent (Boehringer-Mannheim).

the purification of a ribonuclease (RNase) from a commercial hCG preparation. The pure enzyme killed KS cells in vitro, apparently by apoptosis, suggesting that effects of commercial hCG preparations on KS cells and tumours are due, at least in part, to RNase present as a contaminant in these preparations.

We have shown a close association between a human urinary RNase (apparent relative molecular mass (M_r) 18,000) and the β -core fragment of hCG (ref. 5). During the present study we purified the RNase to electrophoretic homogeneity using gel permeation and ribonuclease inhibitor-affinity chromatographies (Fig. 1a; specific activity 892 units per mg of protein (ref. 6)). After electroblot procedures the pure RNase did not crossreact with monoclonal or polyclonal antibodies raised against hCG \(\beta\)-core protein, but it had potent dose-dependent killing activity against a KS Y-1 cell line1 (Fig. 1b). Furthermore, the RNase induced apoptosis in the KS Y-1 cells in a dose-dependent manner (Fig. 2a). It had no effect on a HeLa cell line (Fig. 1b and data not shown).

Until now, anti-KS activity detected in commercial preparations of chorionic gonadotrophin has been attributed solely to hCG. The present results indicate that an RNase present in these crude preparations makes a major contribution to anti-KS activity. Indeed, the close association of the β-core of hCG with the RNase during purification procedures⁵ might explain why partly purified β-core preparations contain particularly potent anti-KS activity. In this regard, recent results obtained by Albini et al.2 must be borne in mind. These authors found that recombinant hCG inhibited growth of KS cells and that the inhibitory effect was blocked by antihCG antibodies. It therefore seems likely that at least some of the anti-KS activity detected in commercial hormone preparations is attributable to intact or fragmented hCG. Thus, overall potent anti-KS activity of commercial hCG preparations might result from the combined effects of hCG and RNase activities.

Amino-terminal sequence analysis of the 18K RNase present in human urine indicates that the enzyme belongs to the RNase superfamily that includes onconase^{5,7}. Onconase is an amphibian ribonuclease that is cytotoxic to cancer cells in vitro and exhibits anti-tumour activity in animal models⁸; it has shown promise in the treatment of human cancers and is currently in phase III clinical trials in patients with pancreatic carcinoma^{9,10}. The 18K human RNase might have a similar role in protection against neoplastic disease. It may prove possible to exploit the anti-neoplastic properties of this enzyme as part of a new regime for the treatment of AIDS-related KS and other tumours.

Suzanne J. Griffiths, David J. Adams

Department of Microbiology, University of Leeds, Leeds LS2 9JT, UK e-mail: d.j.adams@leeds.ac.uk

Simon I. Talbot

Institute of Cancer Research, 237 Fulham Road,

London, SW3 6JB, UK

- 1. Lunardi-Iskandar, Y. et al. Nature 375, 64-68 (1995).
- Albini, A. et al. AIDS 11, 713-721 (1997).
- 3. Harris, P. J. Lancet 346, 118-119 (1995).
- 4. Gill, P. S. et al. N. Engl. J. Med. 335, 1261-1269 (1996).
- 5. Griffiths, S. J., Bramley, T. A., Menzies, G. S. & Adams, D. J. Mol. Cell. Endocrinol. 134, 69-76 (1997).
- Sakakibara, R., Hashida, K., Kitahara, T. & Ishiguro, M. J. Biochem, (Tokyo) 111, 325-330 (1992).
- 7. Schein, C. H. Nature Biotechnol, 15, 529-536 (1997).
- 8. Mikulski, S. M., Ardelt, W., Shogen, K., Bernstein, E. H. & Menduke, H. J. Natl Cancer Inst. 82, 151-153 (1990).
- Mikulski, S. M., Grossman, A. M., Carter, P. W., Shogen, K. & Costanzi, J. J. Int. J. Oncol. 3, 57-64 (1993).

10. Newton, D. L. et al. Protein Eng. 10, 463-470 (1997).